

Identification of a Neocentromere in a Rearranged Y Chromosome With No Detectable DYZ3 Centromeric Sequence

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An 18-year-old woman was evaluated because of primary amenorrhea and hypogonadism. Chromosome analysis from peripheral blood lymphocytes revealed a nonmosaic 46,X,+mar constitution. The marker was shown to be a rearranged Y chromosome consisting of an inverted duplication of the long arm: rea(Y)(qter-q11::q11-qter). Deletion mapping analysis with Y-specific STS showed that the marker lacked Yp and Y-centromeric (DYZ3) sequences, but it was positive for Yq sequences tested. Fluorescence in situ hybridization analysis with Y and X chromosome centromeric and pancentromeric probes showed no hybridization signals. The marker chromosome is present in 100% of the cells; therefore, it is mitotically stable despite the absence of DYZ3 centromeric sequence. Hybridization with CENP-A and CENP-C specific antibodies localized a neocentromere close to the breakpoint.

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INTRODUCTION

In eukaryotes, the centromere, cytogenetically defined as the primary constriction, is the site of sister chromatid attachment. It is essential for proper segregation of the chromosomes during mitosis and meiosis [Pluta et al., 1995]. The mammalian centromere contains large amounts of highly repeated satellite DNA, the best characterized of which is alpha-satellite DNA [Manuelidis, 1978].

Alpha-satellite DNA is believed to be important for centromeric function because it is the only type of sequence shown to be present at the primary constriction of all human chromosomes [Manuelidis, 1978]. The introduction of artificial chromosomes carrying alpha-satellite DNA into mammalian cells has demonstrated that this DNA sequence provides some, if not all, information required in cis for the formation of the centromere [Haff et al., 1992; Larin et al., 1994; Harrington et al., 1997; Ikeno et al., 1998].

Conflicting evidence for alphoid satellite as an essential DNA component of functional centromeres has emerged from the study of stable marker chromosomes, which fail to show labeling with specific alpha-satellite DNA probes. These alphoid chromosomes carry newly derived centromeres (called "neocentromeres") that are apparently formed within interstitial chromosomal sites that have not previously been known to express centromere function [Choo, 1997; du Sart et al., 1997; Warburton et al., 2000]. Moreover, in human dicentric chromosomes, the alpha-satellite DNA is present on both the active and inactive centromeres, suggesting that the presence of alpha satellite per se is insufficient to determine centromere function [Earnshaw et al., 1989; Warburton, 2001].

Five constitutive centromere-binding proteins have been implicated in centromere function: CENP-A, CENP-B, CENP-C, CENP-G and CENP-H. Three of them (CENP-A, -C and -H) associate specifically with active centromeres, that is, are present on normal

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centromeres and neocentromeres and absent from inactive centromeres [reviewed by Warburton, 2001]. These CENPs may be responsible for the establishment of an epigenetic mark, which determines the propagation of human centromeres at particular chromosome locations.

One effective genetic approach to the learning about mammalian centromeres is to examine structurally rearranged chromosomes with abnormal centromeres. We report on an unusual rearranged Y chromosome with no detectable alphoid DNA bearing a functional neocentromere close to the breakpoint.

PATIENTS AND METHODS

Patient Description

An 18-year-old girl was referred to us due to primary amenorrhea and hypogonadism. She was born at term to nonconsanguineous and healthy parents. She had two healthy sibs (a 10-year-old sister and a 5-year-old brother). Family history was unremarkable. There was a history of a slight delay on motor and speech development, learning difficulties, and nocturnal enuresis. When she was 4 years old, a left inguinal hernia surgery was performed.

On physical examination, there was normal height (162 cm) and weight (64.4 kg), macrocephaly (head circumference = 59 cm), M-shaped nuchal hairline, posterior rotation of the ears, prominent forehead, a diffuse goiter, widely spaced nipples, cubitus valgus, short fourth and fifth metacarpals and fifth metatarsals, high frequency of whorl pattern on fingertips, a right simian crease, and clinodactyly of the fifth toes. There was no breast development, external genitalia were female, gonads were not palpable, and pubic hair was on Tanner stage IV with a female distribution.

Laboratory investigation showed high levels of follicle-stimulating hormone (68.7 mIU/mL) and luteinizing hormone (42.1 mIU/mL), low levels of estradiol (12 pg/mL) and total testosterone (1.5 ng/mL), normal levels of thyroid-stimulating hormone (0.61 μ UI/mL) and free thyroxine (15.74 pmol/L), and negative thyroid peroxidase and thyroglobulin antibodies. Bone age was 14 years, and there was lumbar osteopenia. Echocardiogram showed mitral valve prolapse. Ultrasonography revealed hypoplastic uterus (4.95 cm³) and the gonads were not detected. Bilateral gonadectomy was performed and histology revealed streak gonads.

Cytogenetic and FISH Studies

Cytogenetic analysis was performed on metaphase chromosomes obtained from phytohemagglutinin-stimulated peripheral blood lymphocytes using GTG, CBG and QFQ-banding. One hundred cells were analyzed. Fluorescence in situ hybridization (FISH) to metaphase cells using alpha-DNA satellite probes for X and Y chromosomes (DXZ1 and DYZ3, Oncor, Gaithersburg, MD), X-painting probe (COATSOME⁺ X, Oncor), and human alpha-satellite pancentromere probe (Oncor) were performed according to the manufacturer's instructions. Probe HY10 consists of a 3.4-kb

Y-specific repeat, which is a major component of the Y heterochromatin long arm [Nakahori et al., 1986]. Immunofluorescence using antibodies to CENP-A or CENP-C and simultaneous FISH with probe HY10 were performed essentially as described [Warburton et al., 1997, 2000].

Molecular Studies

DNA was extracted from peripheral blood [Sambrook et al., 1989]. The sequences TSPY and DYZ3 were investigated by polymerase chain reaction (PCR) using external primers as in Binder et al. [1995]. DYZ3 sequence was reamplified in a nested-PCR reaction with internal primers [Binder et al., 1995]. The following Y-specific STS—sy81, sy86, sy151, sy117, sy143, sy254, sy255—were analyzed by PCR as described by Vollrath et al. [1992] and Reijo et al. [1995]. The presence of the SRY gene was investigated with primers XES10 and XES11 [Hawkins et al., 1992]. Each PCR contained normal female and male controls. A female operator performed all reactions, including DNA extraction.

RESULTS

Chromosome analysis from peripheral blood lymphocytes with G-, Q-, and C-banding revealed a nonmosaic 46,X,+mar constitution. The marker consisted of two blocks of Q-positive heterochromatin separated by a region of Q-negative euchromatin. C-banding also revealed positive bands on both marker extremities. The banding patterns suggested that the marker might be an i(Yq). The karyotype of her brother was normal (46,XY), as well as that of her sister (46,XX). Although the father was not available for examination, the presence of a normal Y chromosome in the karyotype of the brother suggests that the formation of this Y rearranged chromosome occurred de novo, assuming same paternity of the sibs.

Molecular techniques were employed to confirm the Y origin of the marker, and a deletion map was established (Fig. 1). PCR studies showed positive results for all Yq sequences tested. However, Yp (SRY, TSPY) and Y centromeric (DYZ3) sequences tested negative. DYZ3, which corresponds to the chromosome Y centromeric alpha-satellite DNA, was assayed by nested PCR and the result was negative. The marker was therefore interpreted as a rea(Y)(qter-q11.2::q11.2-qter).

FISH experiments were carried out to search for possible centromere sequences within this marker. The marker failed to show labeling with X and Y centromeric probes and with X painting probe (data not shown). FISH with a human pancentromeric probe (Oncor) showed hybridization to all normal centromeres but not to the marker (Fig. 2a). CENP-C- and CENP-A-specific antibodies were shown to stain a region at or near the center of the chromosome, close to the breakpoint (Fig. 2b,c).

DISCUSSION

We report in this article a rearranged Y chromosome found in a patient with a 46,X,rea(Y)(qter-q11.2::q11.2-qter) chromosomal constitution. This rearranged Y

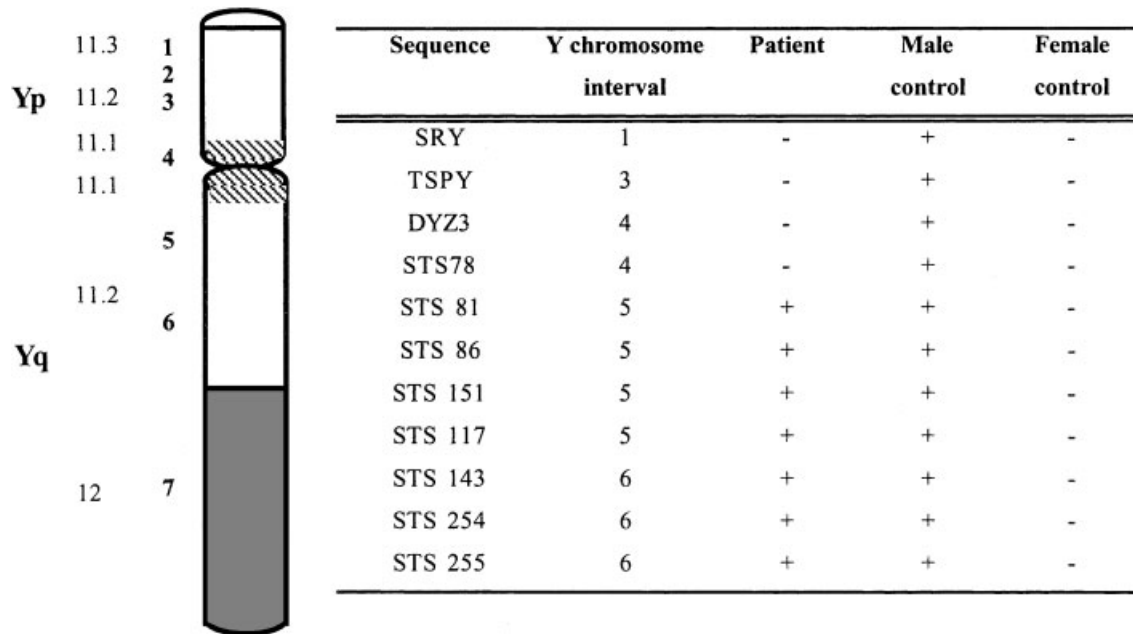


Fig. 1. Molecular map of the rearranged Y chromosome (+, sequence present; -, sequence absent).

chromosome is present in 100% of peripheral blood cells and therefore has been efficiently retained through cell divisions despite the absence of the endogenous centromere region. The presence of a neocentromere on this marker was confirmed by the absence of detectable alpha-satellite DNA and the presence of CENP-A and CENP-C (Fig. 2). This marker chromosome raises the question of the functional requirements for centromere formation.

Four neocentromeres have been reported on derivative Y chromosomes. One previously reported neocentric Y chromosome was similar to the one reported here, although apparently contained slightly less euchromatic material (Fig. 1) [Florida et al., 2000]. This

marker consisted of an inverted duplication of the long arm heterochromatin and a small amount of euchromatin, with deletion of the endogenous Y centromere and alpha-satellite DNA [Florida et al., 2000]. The centromeric protein-binding domain in this marker was shown to be located within the DAZ gene cluster in Yq11.2. This chromosomal region is deleted in some infertile males [Reijo et al., 1995].

Thus, in the two reported cases of inverted duplication neocentric Y chromosome derivatives, the neocentromere appears to be found in the euchromatic DNA at or near the breakpoint [Florida et al., 2000] (Fig. 2b,c). These chromosomes are consistent with the most common mechanism for the formation of anaphoid marker

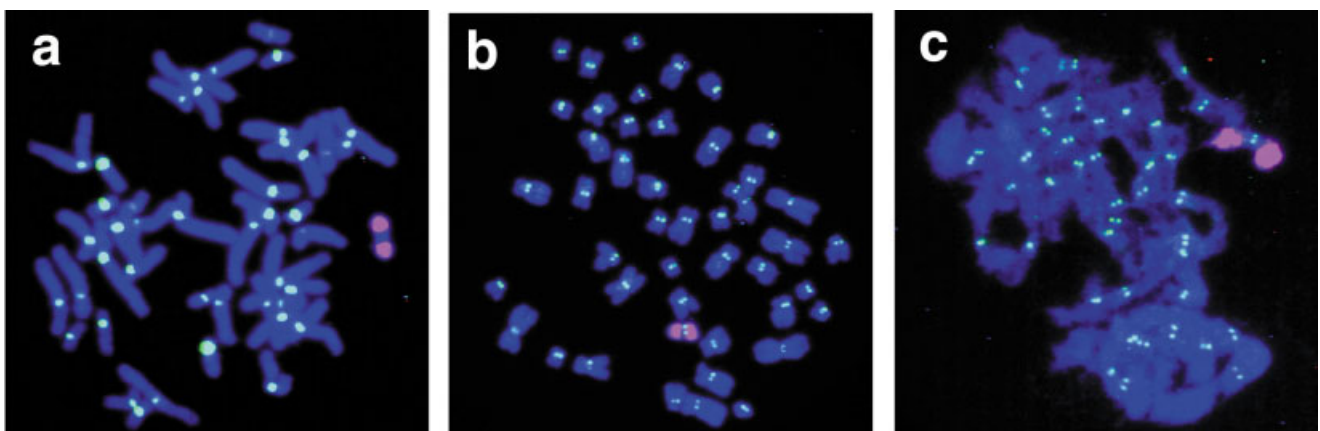


Fig. 2. Identification of alphoid DNA sequences and centromeric proteins. **a**: Fluorescence in situ hybridization (FISH) with a pancentromeric probe is shown in green and the derivative Y chromosome is identified by FISH using probe HY10 in pink. Immunofluorescence using antibodies to CENP-C (**b**) or CENP-A (**c**) is shown in green and the derivative Y chromosome is identified by FISH using probe HY10 in pink. [Color figure can be viewed in the online issue, which is available at www.interscience.wiley.com.]

chromosomes, the de novo inverted duplication of distal segments of chromosomes, resulting in mirror-image chromosomes [Choo, 1997; Warburton et al., 2000]. In the majority of these cases, the neocentromeres are found on one of the duplicated arms, although there are reports of metacentric chromosomes whose neocentromeres are at or near the breakpoints of the inverted duplications [Choo, 1997].

The other three cases of neocentromere-containing Y chromosomes are found on "neodicentric" Y chromosomes, which contain both an inactivated endogenous Y centromere and a neocentromere within or very close to the heterochromatin in the long arm [Bukvic et al., 1996; Rivera et al., 1996; Tyler-Smith et al., 1999]. It is of interest that to date such "neodicentric" chromosomes have only been observed on Y chromosomes. The endogenous Y centromeres may be less stable and more easily inactivated than other centromeres, due to the relatively small arrays of diverged alpha-satellite DNA and lack of binding sites for the heterochromatin protein CENP-B [Tyler-Smith et al., 1999]. Furthermore, the large amounts of constitutive heterochromatin in Y chromosome long arms may predispose formation of neocentromeres. Thus, the Y chromosome may be uniquely suited to form these unusual chromosomes.

The analysis of rearranged Y chromosomes allowed a mitotic centromeric interval to be defined consisting of approximately 150 kb of the alphoid array and about 300 kb of Yp adjacent short arm sequences [Tyler-Smith et al., 1993]. Yeast Artificial Chromosomes (YACs) containing human Y alphoid DNA were introduced into hamster and human cells and re-formed several of the properties of a centromere [Larin et al., 1994]. This suggests that Y alphoid DNA contains the information required to specify some of the centromeric functions. However, human markers with neocentromeres that contain no detectable alpha-satellite DNA indicate that this DNA is not mandatory for centromere function [Choo, 1997; Warburton et al., 2000]. The detailed analysis of a human chromosome 10-derived neocentromere revealed that the neocentromere sequence is not similar to known centromeric sequences. It is possible that the overall composition and distribution patterns of various unknown functional elements, or any "ordinary" DNA under appropriate epigenetic influences, determine centromere formation and function [Barry et al., 1999; Lo et al., 2001].

Hsu [1994] reviewed seven nonmosaic cases of monocentric isochromosomes for the long arm of Y. As expected, these individuals were all phenotypic females with sexual infantilism since they lack the *SRY* gene. Streak gonads were reported in all cases in which this information was available, whereas Turner syndrome (TS) features and short stature were described in at least half of them. Therefore, the female phenotype, the normal height, and the TS features found in our patient are in accordance with her karyotype. It is not clear yet whether the slight delay in motor and speech development observed in childhood has any relation with the chromosomal rearrangement observed. Further characterization of patients with rearranged Y chromosomes might help to clarify these observations.

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REFERENCES

- Barry AE, Howman EV, Cancilla MR, Saffery R, Choo KHA. 1999. Sequence analysis of an 80 kb human neocentromere. *Hum Mol Genet* 8:217–227.
- Binder G, Kich A, Wajs E, Ranke MB. 1995. Nested polymerase chain reaction study of 53 cases with Turner's syndrome: is cytogenetically undetected Y mosaicism common? *J Clin Endocrinol Metab* 80:3532–3535.
- Bukvic N, Susca F, Gentile M, Tangari E, Ianniruberto A, Guanti G. 1996. An unusual dicentric Y chromosome with a functional centromere with no detectable alpha-satellite. *Hum Genet* 97:453–456.
- Choo KHA. 1997. Centromere DNA dynamics: latent centromeres and neocentromeres formation. *Am J Hum Genet* 61:1225–1233.
- du Sart D, Cancilla MR, Earle E, Mao J, Saffery R, Tainton KM, Kalitsis P, Martyn J, Barry AE, Choo KHS. 1997. A functional neo-centromere formed through activation of a latent human centromere and consisting of non-alpha-satellite DNA. *Nature Genet* 16:144–153.
- Earnshaw WC, Ratrie H, Steeten G. 1989. Visualization of centromere proteins CEBP-B and CENP-C on a stable dicentric chromosome in cytologica spreads. *Chromosoma* 98:1–12.
- Florida G, Gimelli G, Zuffardi O, Earnshaw WC, Warburton PE, Tyler-Smith C. 2000. A neocentromere in the *DAZ* region of the human Y chromosome. *Chromosoma* 109:318–327.
- Haff T, Warburton PE, Willard HF. 1992. Integration of human alpha-satellite DNA into simian chromosomes: centromere protein binding and disruption of normal chromosome segregation. *Cell* 70:681–696.
- Harrington JJ, Bokkelen GV, Mays RW, Gustashaw K, Willard HF. 1997. Formation of *de novo* centromeres and construction of first-generation human artificial minichromosomes. *Nature Genet* 15:345–355.
- Hawkins JR, Taylor A, Berta P, Levilliers J, Van der Auwera B, Goodfellow PN. 1992. Mutational analysis of *SRY*: nonsense and missense mutations in XY sex reversal. *Hum Genet* 88:471–474.
- Hsu LYF. 1994. Phenotype/karyotype correlations of Y chromosome aneuploidy with emphasis on structural aberrations in postnatally diagnosed cases. *Am J Med Genet* 53:108–140.
- Ikeno M, Grimes B, Okazaki T, Nakano M, Saitoh K, Hoshino H, McGill NI, Coode H, Masumoto H. 1998. Creation of human artificial chromosomes by introduction of YACs retrofitted with human telomeric DNA. *Nat Biotech* 16:431–439.
- Larin Z, Fricker MD, Tyler-Smith C. 1994. *De novo* formation of several features of a centromere following introduction of a Y alphoid YAC into mammalian cells. *Hum Mol Genet* 3:689–695.
- Lo AW, Craig JM, Saffery R, Kalitsis P, Irvine DV, Earle E, Magliano DJ, Choo KH. 2001. A 330 kb CENP-A binding domain and altered replication timing at a human neocentromere. *EMBO J* 20:2087–2096.
- Manuelidis L. 1978. Chromosomal localisation of complex and simple repeated human DNAs. *Chromosoma* 66:23–32.
- Nakahori Y, Mitani K, Yamada M, Nakagome Y. 1986. A human Y-chromosome specific repeated DNA family (*DYZ1*) consists of a tandem array of pentanucleotides. *Nucl Acids Res* 14:7569–7580.
- Pluta AF, Mackay AM, Ainsztein AM, Goldberg IG, Earnshaw WC. 1995. The centromere: hub of chromosomal activities. *Science* 270:1591–1594.
- Reijo R, Lee TY, Salo P, Alagappan R, Brown LG, Rosenberg M, Rozen S, Jaffe T, Satraus D, Hovatta O, de la Chapelle A, Siber S, Page DC. 1995.

- Diverse spermatogenic defects in humans caused by Y chromosome deletions encompassing a novel RNA-binding protein gene. *Nat Genet* 10:383–393.
- Rivera H, Vasquez AI, Ayala-Madrigal ML, Ramirez-Duenas ML, Davalos IP. 1996. Alphoidless centromere of a familial unstable inverted Y chromosome. *Ann Genet* 39:236–239.
- Sambrook J, Fritsch EF, Maniatis TE. 1989. *Molecular cloning: a laboratory manual*. 2nd edition. New York: Cold Spring Harbor Laboratory, Cold Spring Harbor.
- Tyler-Smith C, Oakey RJ, Larin Z, Fisher RB, Crocker M, Affara NA, Ferguson-Smith MA, Muenke M, Zuffardi O, Jobling MA. 1993. Localization of DNA sequences required for human centromere function through an analysis of rearranged Y chromosomes. *Nature Genet* 5:368–375.
- Tyler-Smith C, Gimelli G, Giglio S, Florida G, Pandya A, Terzoli G, Warburton PE, Earnshaw WC, Zuffardi O. 1999. Transmission of a fully functional human neocentromere through three generations. *Am J Hum Genet* 64:1440–1444.
- Vollrath D, Foote S, Hilton A, Brown LG, Beer-Romero P, Bogan JS, Page DC. 1992. The human Y chromosome: a 43-interval map based on naturally occurring deletions. *Science* 258:52–59.
- Warburton PE. 2001. Epigenetic analysis of kinetochore assembly on variant human centromeres. *TIG* 17:243–247.
- Warburton PE, Cooke CA, Bourassa S, Vafa O, Sullivan B, Stetten G, Gimelli G, Warburton D, Tyler-Smith C, Sullivan KF, Poirier GG, Earnshaw WC. 1997. Immunolocalization of CENP-A, a kinetochore-specific histone H3 variant, suggests a distinct nucleosome structure at the inner kinetochore plate of active centromeres. *Curr Biol* 7:901–904.
- Warburton PE, Dolled M, Mahmood R, Alonso A, Li S, Naritomi K, Tohma T, Nagai T, Hasegawa T, Ohashi H, Govaerts LCP, Eussen BHJ, Hemel JOV, Lozzio C, Schwartz S, Dowhanick-Morissette JJ, Spinner NB, Rivera H, Crolla JA, Yu C, Warburton D. 2000. Molecular cytogenetic analysis of eight inversion duplications of human chromosome 13q that each contain a neocentromere. *Am J Hum Genet* 66:1794–1806.